

Elasticity theory of a twisted stack of plates

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Abstract. We present an elastic model of B-form DNA as a stack of thin, rigid plates or base pairs that are not permitted to deform. The symmetry of DNA and the constraint of plate rigidity limit the number of bulk elastic constants contributing to a macroscopic elasticity theory of DNA to four. We derive an effective twist-stretch energy in terms of the macroscopic stretch ϵ along and relative excess twist σ about the DNA molecular axis. In addition to the bulk stretch and twist moduli found previously, we obtain a twist-stretch modulus with the following remarkable properties: 1) it vanishes when the radius of the helical curve following the geometric center of each plate is zero, 2) it vanishes with the elastic constant K_{23} that couples compression normal to the plates to a shear strain, if the plates are perpendicular to the molecular axis, and 3) it is nonzero if the plates are tilted relative to the molecular axis. This implies that a laminated helical structure *carved* out of an isotropic elastic medium will not twist in response to a stretching force, but an isotropic material will twist if it is *bent* into the shape of a helix.

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1 Introduction

The elastic properties of DNA have become a focus of recent research [1–6]. In particular, stretching experiments on single molecules of DNA now provide a direct probe of the bending, stretching, and twisting elasticity of DNA [7, 8, 10]. Understanding the elasticity of single molecules of DNA may be relevant *in vivo*; *e.g.*, the recA protein which is responsible for homologous recombination in bacterial mitosis has been observed to stretch and twist DNA when it is bound to the DNA molecule [9].

Marko and Siggia recently modeled DNA as a thin, uniform rod with a linear bending elasticity and calculated the extension of the rod as a function of the applied stretching force [2]. This worm-like chain model was appropriate for stretching forces $f \ll \gamma$, where $\gamma \sim Ak_B T/R^2 \approx 200$ pN is the stretch modulus, $A \approx 50$ nm is the bend persistence length, and $R = 1$ nm is the radius of the molecule. Forces in this regime pull on the thermal fluctuations of the molecular backbone but do not pull on the internal structure of DNA. The worm-like chain model accurately predicts the extension of the molecule to within 10% up to approximately 10 pN [7]. Above 10 pN [7, 8] there are no longer thermal effects and DNA is stretched *elastically*. A strain variable ϵ must be introduced to describe increases in the molecular length after thermally

induced contour length fluctuations are fully stretched [2, 11, 7, 8].

DNA is also characterized by the degree of twist of its phosphate backbones about the central axis. Relative twist excess or deficit from a state of preferred twist is parameterized by a signed twist variable σ . The chiral asymmetry of DNA allows for a twist-stretch coupling between σ and ϵ ; this coupling is clearly visible in the recent experiments of Strick *et al.* [10, 4, 5]. Theoretical work has focused on calculating the extension of the molecule as function of both the applied stretching force and the applied excess twist [3, 6, 13]. However, in this paper we investigate a more microscopic origin of the twist-stretch coupling [14].

We introduce a simple elastic model of B-form DNA in which the molecule is viewed as a stack of thin, rigid plates that represent base pairs and are rotated and displaced relative to each other. (See Sect. 3 below for a review of the geometry of B-form DNA.) The centers of mass of the plates define a helical path around a straight central line, which we call the molecular axis. The perpendicular distance from the molecular axis to the helix is called the helix axis offset. Distortion energies in this model are controlled by an underlying continuum elastic energy with those elastic constants allowed by the anisotropic symmetry of DNA. Our model of DNA as a collection of thin, rigid plates has four contributing elastic constants coupling strains with spatial variation normal to the plates.

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Of particular importance is the elastic constant K_{23} coupling compression perpendicular to the plates to a shear displacement parallel to the plates. K_{23} vanishes if the phosphate backbones point in the same direction¹ and also vanishes in the isotropic limit in which each plate has reflection and C_4 or higher symmetries.

We calculate the twist-stretch coupling in terms of its elastic constants and the small helix axis offset. We find that the twist-stretch coupling has terms linear and quadratic in the helix axis offset. The coefficient of the linear term is proportional to the elastic constant K_{23} . In the isotropic limit or when K_{23} vanishes, the twist-stretch coupling is quadratic in the helix axis offset as we calculated earlier [5,12] in related but simpler models.

An important simplification of our model is the constraint that the plates comprising the DNA stack are rigid and undeformable. This is equivalent to setting to infinity all elastic constants coupling the strains within a plate. We thus neglect propeller, buckle, opening, *etc.* deformations [16] of individual base pairs and focus instead on inter-base pair deformations described by relative rotations (roll, tilt, and twist) and translations (slide, shift, and rise) of the base pairs [15,16]. We believe this is a reasonable approximation for stretching forces in the range $1 \text{ pN} < f < 10 \text{ pN}$ and relative twist excesses $|\sigma| < 0.03$.

2 Description of the model

DNA is composed of base pairs connected by two oppositely-directed sugar-phosphate backbones that wrap in two helices around the central molecular axis to produce the major and minor grooves. A cross-section of DNA normal to the phosphate backbones is shown in Figure 1. To construct our model for DNA, we first imagine unwinding the helix so that the phosphate backbones describe two straight, parallel paths. This unwound structure is an elastic rod characterized by an elastic constant tensor K_{ijkl} with components constrained by the symmetry of the rod. Each cross-section of the rod looks like Figure 1. Let the z -axis be parallel to the phosphate backbones, and let the x -axis pass through the midpoint and perpendicular to the line connecting the backbones. The only symmetry operation of this rod is a rotation through π about the x -axis that causes the axes to transform as $x \rightarrow x$, $y \rightarrow -y$, and $z \rightarrow -z$. This symmetry allows for a maximum of 13 independent elastic constants. Our assumption of the rigid shape of each cross-section in the x - y plane eliminates from consideration all strains that require spatial derivatives with respect to x or y . Thus we need only consider the strains u_{zz} , u_{zx} , and u_{zy} and the four elastic constants $K_{33} = K_{zzzz}$, $K_{11} = K_{zzxx}/4$, $K_{22} = K_{zzyy}/4$, and $K_{23} = K_{zyzz}/2$ permitted by symmetry that couple these strains. The elastic constant K_{23} is allowed because the two phosphate backbones are oppositely-directed. If they were undirected or pointed in the same direction, the reflection $y \rightarrow -y$ would be a symmetry and K_{23} would be

¹ Each phosphate backbone has a particular orientation: either 3'-5' or 5'-3'.

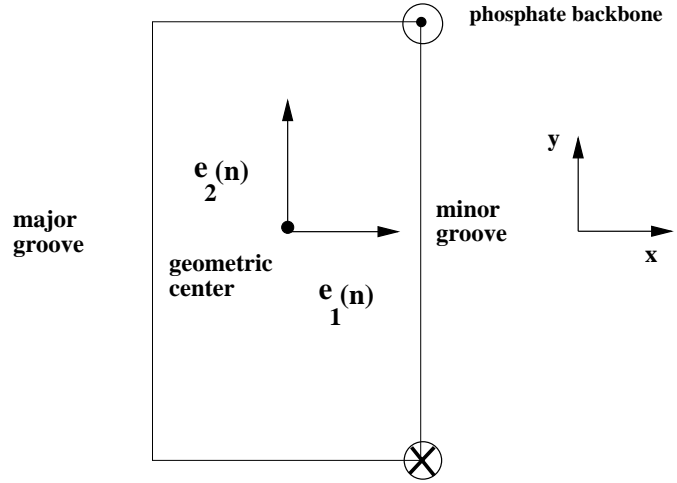


Fig. 1. Schematic cross section of DNA taken perpendicular to the sugar-phosphate backbones. \mathbf{e}_1 is perpendicular to the line connecting the two phosphate backbones and points from the geometric center of the plate toward the minor groove and \mathbf{e}_2 points from the geometric center toward the backbone running in the positive \mathbf{e}_3 direction. Each slice has a coordinate system (x, y, z) with \mathbf{e}_1 as the x -axis, \mathbf{e}_2 as the y -axis, and \mathbf{e}_3 as the z -axis. Also, each slice is labeled by an integer n .

zero. If, in addition, a rotation by $\pi/2$, which transforms $x \rightarrow y$ and $y \rightarrow -x$, were a symmetry, there would be only two elastic constants K_{33} and $K = K_{11} = K_{22}$. We will refer to this case with two elastic constants as the isotropic limit. We view the cross-sectional slices as rigid plates that are coupled elastically *via* the elastic constants K_{33} , K_{11} , K_{22} , and K_{23} .

We now imagine that chiral forces inherent to DNA distort the straight rod to a helical structure in which the centers of mass of the rigid plates describe a helical path about a straight helical axis. This helical structure can be produced by displacing and rotating neighboring plates by a constant amount as depicted in Figure 2. We assume that chiral energies leading to a ground-state helical structure are linear in strain (*i.e.* surface terms) so that the energies of distortions from the ground state are determined by the elasticity of the original untwisted rod, *i.e.*, by the elastic constants K_{33} , K_{11} , K_{22} , and K_{23} . In the following sections, we will derive the twist-stretch coupling in terms of these constants and the helix axis offset.

3 Geometry of B-form DNA

We now consider three different ways of stacking our thin, rigid plates to create a helical model of the straight state of B-DNA. Three possible configurations of straight B-DNA resulting from the rotations and translations described in Figure 2 are shown in Figure 3. We first imagine simply twisting the plates about the long axis passing through the center of mass of the original rod to create shape I. In this shape, the geometric centers of each slice lie on top of one another, the slices are rotated about the long axis relative to one another, and the slices remain perpendicular to

the long axis. Shape I is shown in Figure 3a. Shape II is obtained by first twisting the plates about the long axis and then removing material from the side of each plate opposite to the phosphate backbones. The line connecting the new geometric centers of each slice follows a helix. The slices remain perpendicular to the helix axis about which the geometric centers of each plate rotate. (In the discussion below we use the terms helix axis and DNA molecular axis interchangeably.) Shape II is pictured in Figure 3b. Shape III is obtained by bending the long axis of the sliced rod into a helix. The curve \mathbf{r} connecting the geometric centers of each plate again follows a helix, but now the plates are perpendicular to the local tangent to \mathbf{r} instead of the helix axis. Shape III is shown in Figure 3c.

We note that in the unstressed state of B-form DNA the plates are roughly perpendicular to the molecular axis, and their geometric centers nearly coincide with the molecular axis [17]. Thus to study small deformations of B-DNA, it suffices to restrict our attention to the case where shapes II and III are small perturbations of shape I. In shape II, the amount of material removed from each slice and hence the radius of the helix is small and in shape III the bend of the rod axis away from the molecular axis is also small. Shapes II and III can be described as twisted stacks of plates with small helix axis offsets. If the collection of plates has a helix axis offset, the plates are not stacked with one directly on top of the other; instead, the shift vector (which locates the center of mass of the next plate) has a component perpendicular to the molecular axis. We will see that in our model the helix axis offset is the origin of the twist-stretch coupling.

4 Elasticity theory

In what follows we will develop a long-wavelength elasticity theory for a collection of stacked, rigid plates in terms of small deviations of the shifts and rotations away from their unstressed values. We will then eliminate the shift and rotation variables in favor of the stretch along the molecular axis ϵ and the relative excess twist about the molecular axis σ . The stretch ϵ and relative excess twist σ are the strain variables which have been measured in recent experiments [10,11]. This will allow us to find the bulk twist and stretch moduli in terms of four elastic constants and geometric properties of the molecule. More importantly, it will also enable us to calculate the twist-stretch coupling and determine how it scales with the small helix axis offset.

As shown in Figure 1 we inscribe on each plate a right-handed, orthonormal triad $\mathbf{e}_\alpha(n)$ where $\alpha = 1, 2, 3$ and n is a unitless parameter which labels the plate. In the following, Roman indices run over Cartesian space coordinates (x, y, z) and Greek indices label the plate-fixed coordinate system $(1, 2, 3)$. \mathbf{e}_1 and \mathbf{e}_2 lie in the plane of the plate, and \mathbf{e}_3 is perpendicular to the plate. The coordinates on each plate are labeled by the pair (η_1, η_2) which corresponds to the point on the plate $\eta_1 \mathbf{e}_1(n) + \eta_2 \mathbf{e}_2(n)$. We must also specify the origin on each plate. We choose the origin to be the geometric center (*i.e.*, center of mass) of the plate,

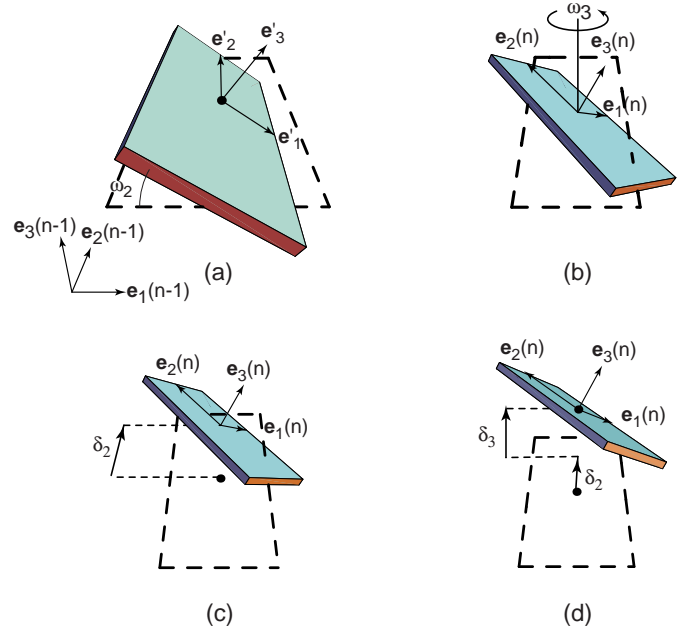


Fig. 2. The orientation and location of a given plate n is generated from the orientation and location of plate $n-1$ by specifying the vector of rotation rates $\boldsymbol{\omega} = (0, \omega_2, \omega_3)$ and the vector of shifts $\boldsymbol{\delta} = (0, \delta_2, \delta_3)$. To obtain the orientation and location of plate n we must apply four operations (two rotations and two shifts) to plate $n-1$. The dashed rectangle shows the outline of plate $n-1$. Note that the rotations ω_i are counterclockwise about $\mathbf{e}_i(n-1)$. (a) We first rotate by ω_2 about $\mathbf{e}_2(n-1)$ and generate a new triad \mathbf{e}'_i . (b) We next rotate the triad \mathbf{e}'_i by ω_3 about $\mathbf{e}_3(n-1)$ and obtain a new triad $\mathbf{e}_i(n)$, the triad at plate n . In (c) we shift the center of plate $n-1$ by δ_2 along $\mathbf{e}_2(n-1)$ and in (d) shift it by δ_3 along $\mathbf{e}_3(n-1)$. This final picture gives the location of the center of plate n and the orientation of its triad.

but this choice is arbitrary and does not affect the elasticity theory, as we will discuss in the Appendix. We can now describe the trajectory in space of the plates and the associated plate-fixed triads. We allow two sets of parameters: a vector of rotation rates $\boldsymbol{\omega}$ and shifts $\boldsymbol{\delta}$. We decompose $\boldsymbol{\omega}$ and $\boldsymbol{\delta}$ in the plate-fixed basis $\mathbf{e}_\alpha(n)$ and assume that the components ω_α and δ_α are constants independent of n .

$$\begin{aligned} \boldsymbol{\omega} &= \omega_\alpha \mathbf{e}_\alpha(n) \\ \boldsymbol{\delta} &= \delta_\alpha \mathbf{e}_\alpha(n), \end{aligned} \quad \text{where } \alpha = 1, 2, 3. \quad (1)$$

The vector of rotation rates $\boldsymbol{\omega}$ describes the rate of change of the orientation of neighboring plates, and thus

$$\frac{d\mathbf{e}_\alpha(n)}{dn} = \boldsymbol{\omega} \times \mathbf{e}_\alpha(n) = -\epsilon_{\alpha\beta\gamma} \omega_\beta \mathbf{e}_\gamma(n), \quad (2)$$

where $\epsilon_{\alpha\beta\gamma}$ is the antisymmetric tensor. The second set of parameters $\boldsymbol{\delta}$ describes the relative displacement of two neighboring plates. We choose $\boldsymbol{\delta}$ to be the relative displacement of the *origin* $\mathbf{r}(n)$ of the (η_1, η_2) coordinate system. We have

$$\frac{d\mathbf{r}(n)}{dn} = \boldsymbol{\delta} = \delta_\alpha \mathbf{e}_\alpha(n). \quad (3)$$

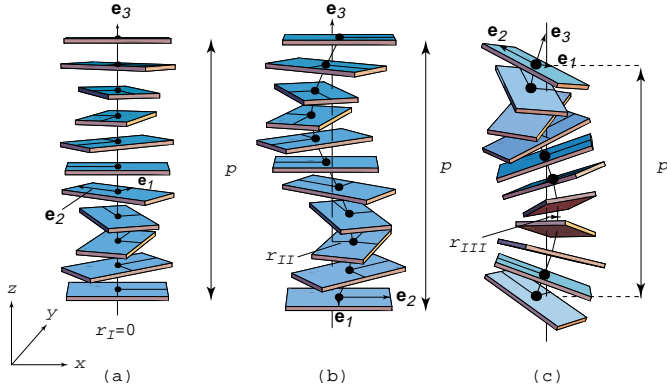


Fig. 3. Three helical shapes obtained from three different stacking rules. The radius and pitch of the helical curves connecting the centers of the plates are given by r and p . (a) Shape I has $\boldsymbol{\delta} = (0, 0, \delta_3)$ and $\boldsymbol{\omega} = (0, 0, \omega_3)$. The plate normal \mathbf{e}_3 is always parallel to the space-fixed z -axis. (b) Shape II has $\boldsymbol{\delta} = (0, \delta_2, \delta_3)$ and $\boldsymbol{\omega} = (0, 0, \omega_3)$. Again, \mathbf{e}_3 is parallel to the z -axis. These vectors generate a helical stack with all of the plates parallel to one another and the centers of the plates offset from the central helix axis by r_{II} . (c) Shape III is generated by $\boldsymbol{\delta} = (0, 0, \delta_3)$ and $\boldsymbol{\omega} = (0, \omega_2, \omega_3)$. The \mathbf{e}_3 axis is now tangent to the helical curve of radius r_{III} defined by the plate centers. The first plate is aligned so that the helix axis is in the vertical direction.

Note that according to our definitions $\boldsymbol{\omega}$ is unitless and $\boldsymbol{\delta}$ has units of length. Finally, the position in space of the point (η_1, η_2) on plate n is simply $\mathbf{x}(\eta_1, \eta_2, n) = \mathbf{r}(n) + \eta_1 \mathbf{e}_1(n) + \eta_2 \mathbf{e}_2(n)$. For constant ω_α and δ_α , these equations will, in general, describe a helical structure. We may solve (2) and (3) to find:

$$\mathbf{r}(n) = \mathbf{e}_\beta(0) \left\{ n \frac{\omega_\beta (\boldsymbol{\delta} \cdot \boldsymbol{\omega})}{\omega^2} + \left(\delta_\beta - \omega_\beta \frac{(\boldsymbol{\delta} \cdot \boldsymbol{\omega})}{\omega^2} \right) \right. \\ \left. \times \frac{\sin(|\boldsymbol{\omega}|n)}{|\boldsymbol{\omega}|} + \frac{\epsilon_{\beta\alpha\gamma} \delta_\alpha \omega_\gamma}{\omega^2} \cos(|\boldsymbol{\omega}|n) \right\}, \quad (4)$$

with $\omega^2 = \omega_\alpha \omega_\alpha$. Examination of (4) shows that the geometric center moves on average in the direction $\mathbf{e}_\beta(0) \omega_\beta / |\boldsymbol{\omega}|$ and traces out a helix with helix axis offset $r = |\boldsymbol{\omega} \times \boldsymbol{\delta}| / \omega^2$ and pitch $p = 2\pi \boldsymbol{\delta} \cdot \boldsymbol{\omega} / \omega^2$. The shift vector, rotation vector, helix axis offset, and pitch are listed in Table 1 for each of the three shapes of B-form DNA we are considering. In Figure 2 we apply the rotation vector $\boldsymbol{\omega} = (0, \omega_2, \omega_3)$ and shift vector $\boldsymbol{\delta} = (0, \delta_2, \delta_3)$ to plate $n - 1$ and obtain the location and orientation of plate n . Successive plates in Figure 2 are constructed by rotating about and shifting along the axes of plates $n, n + 1, n + 2$, *etc.* The three helical stacks discussed in Table 1 are pictured in Figure 3.

In a deformed state, the displacement and rotation rates change to $\boldsymbol{\delta}' = \boldsymbol{\delta} + \boldsymbol{\Delta}(n)$ and $\boldsymbol{\omega}' = \boldsymbol{\omega} + \boldsymbol{\Omega}(n)$ and define new positions $\mathbf{x}'(\eta_1, \eta_2, n)$ for points on plate n . Our goal is to calculate the energy of this deformed state relative to the equilibrium helical state to second order in the small parameters $\boldsymbol{\Delta}$ and $\boldsymbol{\Omega}$, which can in general depend

Table 1. Geometric properties of several helices.

Shape	$\boldsymbol{\delta}$	$\boldsymbol{\omega}$	Helix axis offset	Pitch/ 2π
I	$(0, 0, \delta_3)$	$(0, 0, \omega_3)$	0	δ_3 / ω_3
II	$(0, \delta_2, \delta_3)$	$(0, 0, \omega_3)$	δ_2 / ω_3	δ_3 / ω_3
III	$(0, 0, \delta_3)$	$(0, \omega_2, \omega_3)$	$\omega_2 \delta_3 / (\omega_2^2 + \omega_3^2)$	$\delta_3 \omega_3 / (\omega_2^2 + \omega_3^2)$

on n . We consider plates of thickness δn and calculate the energy in the deformed state of each plate to order δn , ignoring terms of order $(\delta n)^2$ or higher. We then sum over all plates to obtain the total energy of the rod. Since the energy of each plate is proportional to δn , the sum over all plates can be converted into an integral over n .

The energy of the n th plate will depend on the values of $\boldsymbol{\Delta}$ and $\boldsymbol{\Omega}$ at n and, to the order we consider, *not* on their derivatives with respect to n . The energy of each plate will be the same function of $\boldsymbol{\Delta}$ and $\boldsymbol{\Omega}$ for every n , so we need only calculate the energy of a single reference plate. Our elastic theory provides us with distortion energies of the reference plate as a function of the strains $u_{ij} = \frac{1}{2}(\partial_i u_j + \partial_j u_i)$ ($i, j = x, y, z$), where $\mathbf{u} = \mathbf{x}' - \mathbf{x}$ is the displacement variable and x, y , and z are the Cartesian coordinates of the reference plate. By a suitable choice of orientation we may take $\mathbf{e}_1 = \hat{\mathbf{x}}$, $\mathbf{e}_2 = \hat{\mathbf{y}}$, and $\mathbf{e}_3 = \hat{\mathbf{z}}$ for the plate of interest. The free energy for a single plate with thickness δz is (in units of $k_B T$):

$$\frac{F_{\text{plate}}}{k_B T} = \frac{1}{2} \int_z^{z+\delta z} \int dx dy dz K_{ijkl} u_{ij} u_{kl}. \quad (5)$$

Our goal is to express this energy as a function of $\boldsymbol{\Delta}$ and $\boldsymbol{\Omega}$ and change integration variables from x, y , and z to η_1, η_2 , and n . The Jacobian relating the two coordinate systems is found by taking η_α and n derivatives of $\mathbf{x}(\eta_1, \eta_2, n)$ and remembering that $\mathbf{e}_\alpha \cdot \mathbf{e}_\beta = \delta_{\alpha\beta}$.

$$dx dy dz = |\delta_3 + \epsilon_{3\beta\gamma} \omega_\beta \eta_\gamma| d\eta_1 d\eta_2 dn. \quad (6)$$

To calculate the strain tensor u_{ij} we must take the derivatives of $\mathbf{u}(\eta_1, \eta_2, n)$ with respect to η_α and n and then relate these coordinates to Cartesian coordinates x, y , and z fixed on the plate. Since the derivatives of u_i with respect to η_α only include terms proportional to δn , the only derivatives in the strain tensor contributing to the total free energy in (5) are those in the n -direction. We therefore only need to calculate $d\mathbf{u}/dn$. Using the equations of motion, (2, 3), and the expression for $\mathbf{x}(\eta_1, \eta_2, n)$, we have to linear order in Ω_α and Δ_α

$$\frac{d\mathbf{u}(\eta_1, \eta_2, n)}{dn} = \mathbf{e}_\alpha \left[\Delta_\alpha + \epsilon_{\alpha\beta\gamma} \Omega_\beta \eta_\gamma \right] \\ + (\delta_\alpha + \epsilon_{\alpha\beta\gamma} \omega_\beta \eta_\gamma) \left[\mathbf{e}'_\alpha - \mathbf{e}_\alpha \right]. \quad (7)$$

We can neglect the last terms in (7) since they also are proportional to δn . (We note that deformations of the base plates can be included if we retain these δn contributions.)

We choose the slab at n to have its internal triad point along a Cartesian coordinate system (x, y, z) fixed

on the plate with the z -direction aligned with \mathbf{e}_3 and the x - and y -directions aligned with \mathbf{e}_1 and \mathbf{e}_2 respectively. To leading order in Ω_α and Δ_α , the only relevant components of the strain tensor come from the derivatives

$$\partial_z u_i = \frac{\partial u_i}{\partial n} \frac{\partial n}{\partial z} = (\Delta_i + \epsilon_{ijk} \Omega_j \eta_k) / \delta_3, \quad (8)$$

where $\partial n / \partial z = 1 / \delta_3$ for the three shapes we are considering. In (8) the sums over α, β , and γ have been replaced by sums over i, j , and k because we chose the body-fixed coordinates to be along (x, y, z) . We have now reduced the number of effective elastic constants from 13 to 4 since only z derivatives contribute to the free energy. The free energy of a single plate in terms of the strains $\mathbf{\Delta}$ and $\mathbf{\Omega}$ and the coordinates η_1, η_2 , and n is:

$$\frac{F_{\text{plate}}}{k_B T} = \int_n^{n+\delta n} dn \int d^2 \eta \frac{|\delta_3 + \epsilon_{3\beta\gamma} \omega_\beta \eta_\gamma|}{2\delta_3^2} \times \left\{ K_{ij} (\Delta_i + \epsilon_{ilm} \Omega_l \eta_m) (\Delta_j + \epsilon_{jkn} \Omega_k \eta_n) \right\}, \quad (9)$$

where K_{ij} is related to K_{ijkl} by

$$K_{ij} = \begin{cases} \frac{1}{4} K_{zizj} & i = j = x \text{ or } i = j = y \\ \frac{1}{2} K_{zizj} & i = y, j = z \\ K_{zizj} & i = j = z. \end{cases} \quad (10)$$

We see that our elasticity theory of DNA as a stack of thin, rigid plates has four contributing elastic constants, K_{xx} , K_{yy} , K_{yz} , and K_{zz} . Below we will refer to these elastic constants as K_{11} , K_{22} , K_{23} , and K_{33} , respectively, since the plate-fixed axes rotate in space as we move along the helical stack. We note that an isotropic theory has only two contributing constants, $K = K_{11}/2 = K_{22}/2$ and K_{33} . The total free energy for the collection of plates is obtained by adding up the contributions to the free energy from each plate. The total free energy will have the same form as (9) with the Cartesian indices on the strains (x, y, z) replaced by the plate-fixed indices $(1, 2, 3)$.

5 DNA effective free energy

We now sum up the contributions from all slices and do the integrals over η_1 and η_2 in (9) to get an effective theory for the DNA molecule. Before we actually do this, we will make some further simplifications and approximations. We first exploit the symmetry of each slice of the DNA molecule. In Figure 1 we see that rotating each slice by π about \mathbf{e}_1 is a symmetry. Under this symmetry operation η_2 switches sign and hence all moments odd in η_2 are zero: $\langle \eta_2 \rangle = \langle \eta_1^m \eta_2 \rangle = 0$ ($m = 1, 2, 3, \dots$) where $\langle \cdot \rangle \equiv \mathcal{A}^{-1} \int d^2 \eta (\cdot)$ and \mathcal{A} is the cross sectional area in the $\eta_1 - \eta_2$ plane. We also locate the origin of each plate at its geometric center so that $\langle \eta_1 \rangle = 0$.

In the unstressed configuration of B-form DNA the base pairs make an angle of close to $\pi/2$ with the molecular axis [17]. We therefore take $\omega_{1,2} p \ll \delta_3$ where $p \approx 34 \text{ \AA}$ is the pitch of the DNA helix and $\delta_3 \approx 3.4 \text{ \AA}$ is the axial rise of the base pairs. In addition, the geometric centers of the plates are roughly located on the molecular axis [17]; we therefore also assume $\delta_{1,2}/\delta_3 \ll 1$. We will characterize the unstressed straight state of B-DNA by two known macroscopic parameters (the axial rise δ_3 and the twist rate of the plates about the molecular axis $\omega_3 = 2\pi\delta_3/p \approx 0.63 \text{ rad}$) and two unknown microscopic parameters (δ_2 and ω_2). In what follows we will assume that δ_2/δ_3 and ω_2/ω_3 are small and work to quadratic order in these quantities.

The effective free energy is given below in units of $k_B T$. We neglect the ω_2 contribution arising from the Jacobian in (9) since we are only interested in the lowest order terms in ω_2/ω_3 . Including this term does not alter our findings below for the twist-stretch coupling:

$$\frac{F_{\text{DNA}}}{k_B T} = (A/2\delta_3) \int dn \left\{ K_{11} \Delta_1^2 + K_{22} \Delta_2^2 + K_{33} \Delta_3^2 + K_{33} \langle \eta_2^2 \rangle \Omega_1^2 + K_{33} \langle \eta_1^2 \rangle \Omega_2^2 + \left(K_{11} \langle \eta_2^2 \rangle + K_{22} \langle \eta_1^2 \rangle \right) \Omega_3^2 - 2K_{23} \langle \eta_1^2 \rangle \Omega_2 \Omega_3 + 2K_{23} \Delta_2 \Delta_3 \right\}. \quad (11)$$

The diagonal terms in (11) give the energy costs for deviations in $\boldsymbol{\delta}$ and $\boldsymbol{\omega}$ from their unstressed values. For example, K_{33} controls the energy cost of changing the translation and rotation of plate n relative to plate $n-1$ (as depicted in Fig. 2) from δ_3 to $\delta'_3 = \delta_3 + \Delta_3$ and from ω_2 to $\omega'_2 = \omega_2 + \Omega_2$ respectively. These deviations in the rotations and shifts will change the overall shape of the helical stack. For example, adding a nonzero Ω_2 to shape I causes the plates to become tilted relative to the helix axis and the helical curve connecting the centers of the plates to have a nonzero radius. The off-diagonal terms in (11) control the coupling between excess shifts and rotations. To lowest order in ω_2/ω_3 , K_{23} only couples excess shifts along \mathbf{e}_2 to those along \mathbf{e}_3 and excess rotations about \mathbf{e}_2 to those about \mathbf{e}_3 .

6 Twist-stretch coupling

Motivated by recent stretching experiments on torsionally constrained single molecules of DNA [10], we rewrite (11) in terms of the relative overtwist σ about the molecular axis and the relative stretch ϵ along the molecular axis. To accomplish this we must first find ϵ and σ in terms of the rotation and shift variables Ω_i and Δ_i . After we incorporate ϵ and σ into (11), we minimize over the remaining unconstrained variables. This gives us the twist-stretch energy in terms of the elastic constants K_{ij} and the geometric parameters of the DNA helix. We can then identify the stretch modulus B , the twist modulus C , and the twist-stretch modulus D . We note that this twist-stretch

energy neglects the effects of thermal fluctuations; these effects have been studied recently [6,13].

We now derive the twist-stretch coupling for the three shapes described in Table 1 by considering the general case $\omega_2, \delta_2 \neq 0$ and then neglecting $\omega_2 \delta_2$ cross terms. The unstressed configuration is therefore $\boldsymbol{\omega} = (0, \omega_2, \omega_3)$ and $\boldsymbol{\delta} = (0, \delta_2, \delta_3)$. Each of the three equilibrium shapes mentioned previously can be obtained from this parameterization, for instance, shape II is generated by taking $\delta_2 \neq 0$ and $\omega_2 = 0$. We define the stretch along the molecular axis as the relative deviation in the extension L along the molecular axis from its unstressed value L_0 ,

$$\epsilon = \frac{L}{L_0} - 1. \quad (12)$$

To find ϵ in terms of Δ_i and Ω_i we must determine the extension L from (4). We see that each base pair step increases the extension by an amount $\boldsymbol{\delta} \cdot \boldsymbol{\omega}/|\boldsymbol{\omega}|$ in the direction $\boldsymbol{\omega}/|\boldsymbol{\omega}|$. The total extension for N steps in the distorted configuration is therefore

$$L = N \boldsymbol{\delta}' \cdot \frac{\boldsymbol{\omega}'}{|\boldsymbol{\omega}'|} \quad (13)$$

where the primes refer to the distorted state, *i.e.* $\boldsymbol{\delta}' = (\Delta_1, \delta_2 + \Delta_2, \delta_3 + \Delta_3)$. We can now expand (12) to linear order in the Ω_i and Δ_i and to quadratic order in the small parameters δ_2/δ_3 and ω_2/ω_3 :

$$\epsilon = \frac{\Delta_3}{\delta_3} + \left(\frac{\delta_2}{\delta_3} - \frac{\omega_2}{\omega_3} \right) \frac{\Omega_2}{\omega_3} + \frac{\omega_2}{\omega_3} \frac{\Delta_2}{\delta_3} + \frac{\omega_2^2}{\omega_3^2} \frac{\Omega_3}{\omega_3}. \quad (14)$$

Note that when the helix axis offset of the helical stack is zero ($\delta_2 = \omega_2 = 0$), the stretch reduces to $\epsilon = \Delta_3/\delta_3$. We also note that when $\omega_2 = 0$, Δ_3 and Ω_2 are the only distortions contributing to the stretch.

We must express the relative excess link $\sigma = (Lk/Lk_0) - 1$ in terms of the Ω_i , where the linking number Lk is the number of times the plates rotate about the molecular axis over the length of the stack. In its unstressed state, DNA has linking number $Lk_0 = L_0/p$. (We are working in the force regime where the molecular axis is nearly straight, and hence link and twist about the molecular axis are synonymous.) We see from (4) that it takes $2\pi/|\boldsymbol{\omega}|$ steps to complete one revolution about the molecular axis. Therefore in N steps the top plate has rotated by $N|\boldsymbol{\omega}|/2\pi$ relative to the bottom plate. The relative excess twist about the molecular axis is therefore

$$\sigma = \frac{|\boldsymbol{\omega}'|}{|\boldsymbol{\omega}|} - 1, \quad (15)$$

where $\boldsymbol{\omega}' = (\Omega_1, \omega_2 + \Omega_2, \omega_3 + \Omega_3)$. Expanding (15) to linear order in the Ω_i and to quadratic order in ω_2/ω_3 , we find

$$\sigma = \frac{\omega_2}{\omega_3} \frac{\Omega_2}{\omega_3} + \left(1 - \frac{\omega_2^2}{\omega_3^2} \right) \frac{\Omega_3}{\omega_3}. \quad (16)$$

Note that when $\omega_2 = 0$, the base pairs are perpendicular to the molecular axis and the relative excess twist is simply Ω_3/ω_3 .

Solving (14) for Δ_3 and (16) for Ω_3 , we substitute into (11) and minimize over $\Delta_{1,2}$ and $\Omega_{1,2}$. This gives the effective twist-stretch energy per length along the molecular axis:

$$\frac{F_{\text{TS}}(\epsilon, \sigma)}{N \delta_3 k_B T} = \frac{\mathcal{A}}{2} \left(B \epsilon^2 + C \sigma^2 + 2D \epsilon \sigma \right), \quad (17)$$

where N is the total number of plates, B is the stretch modulus, C is the twist modulus, and D is the twist-stretch modulus. To zeroth order in the small parameters δ_2/δ_3 and ω_2/ω_3 the stretch and twist moduli are:

$$B = K_{33} - K_{23}^2/K_{22} \\ C = (\omega_3^2/\delta_3^2) \left(K_{11} \langle \eta_2^2 \rangle + \langle \eta_1^2 \rangle (K_{22} - K_{23}^2/K_{33}) \right). \quad (18)$$

B , C , and D have dimension L^{-3} and therefore the coefficients of the quadratic dimensionless strains in (17) scale as a persistence length divided by the square of the helical pitch as found previously [5]. We can estimate the values of the elastic constants K_{33} and $K_{11} = K_{22} = K/2$ in the isotropic limit using the known values of the stretch and twist persistence lengths and geometric properties of the DNA helix [5]. In this limit $K_{33} = (2\pi)^2 \bar{B}/(p^2 \mathcal{A}) \approx 85 \text{ nm}^{-3}$ and $K = \bar{C}/\mathcal{A}^2 \approx 10 \text{ nm}^{-3}$ where $\bar{B} \approx 80 \text{ nm}$ is the stretch persistence length and $\bar{C} \approx 100 \text{ nm}$ is the twist persistence length [13].

The twist-stretch couplings D_i for the three shapes given in Table 1 are shown below to lowest order in the helix axis offsets r_i :

$$D_I = 0 \\ D_{\text{II}} \approx \frac{K_{23}}{K_{33}} \left(K_{33} - \frac{K_{23}^2}{K_{22}} \right) \frac{\omega_3 r_{\text{II}}}{\delta_3} \\ D_{\text{III}} \approx \frac{K_{23}}{K_{33}} \left(K_{33} - \frac{K_{23}^2}{K_{22}} \right) \frac{\omega_3 r_{\text{III}}}{\delta_3} \\ + \left(K_{11} \frac{\langle \eta_2^2 \rangle}{\langle \eta_1^2 \rangle} + K_{22} - K_{33} \right) \left(\frac{\omega_3 r_{\text{III}}}{\delta_3} \right)^2, \quad (19)$$

where the helix axis offsets for each shape are $r_I = 0$, $r_{\text{II}} = \delta_2/\omega_3$, and $r_{\text{III}} \approx \omega_2 \delta_3/\omega_3^2$. (The twist-stretch modulus $D \approx 10 \text{ nm}^{-3}$ measured in the recent experiment of Strick *et al.* [10] provides an estimate of $K_{23} r$.) In performing the calculation of the twist-stretch moduli we have made two simplifications: 1) we have dropped the quadratic terms in the helix axis offset that are proportional to K_{23} and 2) we have not included the ω_2 contributions arising from the Jacobian in (9). This last simplification does not affect our conclusions concerning the twist-stretch coupling (TS) shown in Table 2 since ω_2 contributions can only affect the twist-stretch coupling of shape III and shape III already has twist-stretch couplings in both the isotropic and anisotropic models.

We see from (19) that the twist-stretch coupling in our rigid plate model vanishes when the helix axis offset r vanishes. However, calculations for a pretwisted rod of material with a three-dimensional isotropic elasticity yield a twist-stretch coupling for zero helix axis offset provided

Table 2. Determination of the twist-stretch coupling for each shape of B-form DNA is shown below. The symbol TS signifies that there is a nonzero twist-stretch coupling.

Shape	Helix Axis Offset	Isotropic Model ($K_{23} = 0$)	Anisotropic Model ($K_{23} \neq 0$)
I	0	0	0
II	δ_2/ω_3	0	TS
III	$\omega_2\delta_3/\omega_3^2$	TS	TS

the rod cross-section is not circular [18]. To achieve consistency with our results, it seems likely that the magnitude of this coupling decreases as the elastic constants coupling in-plane strains (*i.e.*, u_{xx} , u_{xy} , u_{yy}) increase. We can thus view our calculation as the limit when these elastic constants diverge.

We also see that in the isotropic limit ($K_{23} = 0$) the twist-stretch coupling for shape III scales quadratically with the small helix axis offset as found previously in references [5, 12]. However, when the K_{23} anisotropy is nonzero, the twist-stretch coupling scales linearly with the small helix axis offset. We note that the twist-stretch coupling in the isotropic limit vanishes for shape II since it has $\omega_2 = 0$. However, shape III yields a twist-stretch coupling in both the isotropic and anisotropic cases since it has $\omega_2 \neq 0$. This may be explained by considering the manner in which the plates are stacked in the two different geometries (see Fig. 3). In shape II the plates are stacked perpendicular to the molecular axis. In this case, the plates can move along the molecular axis without rotating in response to the stretching force. In shape III the plates are not perpendicular to the molecular axis, but are instead perpendicular to the local tangent $d\mathbf{r}/dn$. In order to align the plates perpendicular to the molecular axis, they must twist. This implies that an isotropic rod *bent* into the shape of a helix will have a twist-stretch coupling, but carving a helical shape out of isotropic material will not produce a twist-stretch coupling.

7 Comparison to the ribbon model of DNA

We compare our plate model to a ribbon model of DNA recently discussed in reference [12]. There we modeled DNA as a thin helical ribbon that is only allowed to stretch along the ribbon axis. Changes in extension along the molecular axis arose from changes in the angle the ribbon tangent makes with the molecular axis and from stretching the ribbon axis. We showed that the twist-stretch coupling scaled quadratically with the helix axis offset if the twist-bend, twist-stretch, and bend-stretch couplings for twists and bends about and stretches along the *ribbon axis* were set to zero.

To make a direct comparison between the two models we consider the unstressed configuration $\boldsymbol{\delta} = (0, 0, \delta_3)$ and $\boldsymbol{\omega} = (0, \omega_2, \omega_3)$ and do not allow stretch along the 1 and 2 axes of each plate by setting $\Delta_1 = \Delta_2 = 0$. This

corresponds to a helical stack of plates with the planes of the plates perpendicular to the tangent to the curve \mathbf{r} connecting the geometric centers of the plates and stretching only allowed along the tangent $d\mathbf{r}/dn$. When we set $\Delta_1 = \Delta_2 = 0$ in (11), we find a free energy identical to the one studied previously [12]. We see below that our expression for the free energy admits all of the couplings found in the ribbon model, namely the bend-stretch ($\Omega_2\Delta_3$), twist-bend ($\Omega_2\Omega_3$), and twist-stretch ($\Omega_3\Delta_3$) couplings. (We have included the ω_2 contributions from the Jacobian and kept each coupling to lowest order in ω_2/ω_3 .)

$$\begin{aligned} \frac{F_{\text{ribbon}}}{k_B T} &= (A/2\delta_3) \int dn \left\{ K_{33} \langle \eta_2^2 \rangle \Omega_1^2 + K_{33} \langle \eta_1^2 \rangle \Omega_2^2 \right. \\ &+ \left(K_{11} \langle \eta_2^2 \rangle + K_{22} \langle \eta_1^2 \rangle \right) \Omega_3^2 + K_{33} \Delta_3^2 \\ &+ 2K_{33} \frac{\omega_2}{\delta_3} \langle \eta_1^2 \rangle \Omega_2 \Delta_3 - 2K_{23} \langle \eta_1^2 \rangle \Omega_2 \Omega_3 \\ &\left. - 2K_{23} \frac{\omega_2}{\delta_3} \langle \eta_1^2 \rangle \Omega_3 \Delta_3 \right\}. \end{aligned} \quad (20)$$

We find that the presence of the the last two terms leads to twist-stretch couplings that scale linearly with the helix axis offset $D \sim (K_{23}/K_{33})\omega_3 r_{\text{III}}/\delta_3$. If we set $K_{23} = 0$, the lowest order contribution is quadratic in the helix axis offset and we obtain the result $D \sim K_{33}(\omega_3 r_{\text{III}}/\delta_3)^2$ found previously [5, 12].

8 Conclusion

We have presented an elasticity theory for B-form DNA modeled as a stack of thin, rigid plates. The symmetry of the DNA molecule and the assumption of plate rigidity enabled us to reduce the number of contributing elastic constants to four. We then derived an effective twist-stretch energy for DNA in terms of the relative excess twist σ about the molecular axis and the stretch ϵ along the molecular axis. We found an effective twist-stretch coupling with terms that scale linearly and quadratically with the small helix axis offset. We have argued that deformations of the base pairs are more costly than deviations in the shifts and rotations of the base pairs from their unstressed values. We therefore conclude that in the force regime where deformations of the plates are negligible, the twist-stretch coupling is due to a nonzero helix axis offset. We have also found that shape II, in which the normals to the base pairs parallel to the molecular axis, does not have a twist-stretch coupling in the limit $K_{23} = 0$. Since recent experiments show that there is a large twist response to stretch, this implies that either the coupling K_{23} is relevant for an elastic description of B-DNA or that the unstressed straight state of B-DNA is composed of base pairs that tilt relative to the molecular axis.

An important next step is to estimate the elastic constant K_{23} to determine the relative magnitudes of the isotropic and anisotropic terms in the twist-stretch coupling. One might also calculate the twist-stretch coupling

for DNA plasmids where the unstressed state of the molecular backbone is circular rather than linear. This calculation could provide a theoretical estimate of the twist-stretch coupling found in experiments on complexes of Rec-A protein with DNA plasmid [9].

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Appendix: Translational invariance of the plate origin

We comment here on the invariance of our model under the choice of the plate origin. This is an important feature since a twisted stack of plates can be described by an infinite number of one-dimensional curves depending on the choice of the reference point on each plate. We proceed by considering a change in the origin, or, in particular a change in the coordinate system such that $\mathbf{x} = (\eta_1, \eta_2, s) \rightarrow \tilde{\mathbf{x}} = (\tilde{\eta}_1, \tilde{\eta}_2, s) + \mathbf{R}$ where \mathbf{R} is an arbitrary vector. While this translation does not affect the relative rotation of two consecutive plates $\boldsymbol{\omega}$, it does affect the *relative translation*. Consider the action of $g = (\boldsymbol{\omega}, \boldsymbol{\delta})$ on \mathbf{x} :

$$\mathbf{x} \xrightarrow{g} \mathbf{x} + \boldsymbol{\delta} - \boldsymbol{\omega} \times \mathbf{x}. \quad (21)$$

We now consider the transformation in (21) shifted by \mathbf{R} and compare it to the transformation $\tilde{g} = (\tilde{\boldsymbol{\omega}}, \tilde{\boldsymbol{\delta}})$ written in terms of the new coordinate system $\tilde{\mathbf{x}}$.

$$\begin{aligned} \mathbf{x} + \mathbf{R} &= \tilde{\mathbf{x}} + \tilde{\boldsymbol{\delta}} - \tilde{\boldsymbol{\omega}} \times \tilde{\mathbf{x}} \\ &\equiv \left[\left(\tilde{\mathbf{x}} - \mathbf{R} \right) + \boldsymbol{\delta} - \boldsymbol{\omega} \times \left(\tilde{\mathbf{x}} - \mathbf{R} \right) \right] + \mathbf{R}. \end{aligned} \quad (22)$$

In order for g and \tilde{g} to act the same way, we must choose $\tilde{\boldsymbol{\omega}} = \boldsymbol{\omega}$ and $\tilde{\boldsymbol{\delta}} = \boldsymbol{\delta} - \boldsymbol{\omega} \times \mathbf{R}$. We note that not only does $(\boldsymbol{\omega}, \boldsymbol{\delta})$ transform as above, but in addition, due to the linearity of the transformation, $(\boldsymbol{\Omega}, \boldsymbol{\Delta})$ transforms the same way. As a result, we see that the derivatives of interest in (8) transform as

$$\begin{aligned} \partial_z u_i &= [\Delta_i - \epsilon_{ijk} \Omega_j \eta_k] / \delta_3 \\ &\longrightarrow \left[\Delta_i + \epsilon_{ijk} \Omega_j \left(\eta_k - R_k \right) + \epsilon_{ijk} \Omega_j \mathbf{R}_k^{\text{tr}} \right] / \delta_3, \end{aligned} \quad (23)$$

where \mathbf{R}_k^{tr} is the projection of \mathbf{R} onto the $\eta_1 - \eta_2$ plane. Since changing the base point on each slab only requires

components of \mathbf{R} in that plane, we see that the last two terms in (23) cancel and the relevant derivatives are invariant with respect to the choice of the origin on the plate. Thus we are free to choose the most convenient origin for any calculation of interest.

References

1. J.F. Marko, E.D. Siggia, *Macromolecules* **24**, 981-988 (1994).
2. J.F. Marko, E.D. Siggia, *Macromolecules* **28**, 8759-8770 (1995).
3. B. Fain, J. Rudnick, *Phys. Rev. E* **55**, 7364-7368 (1997).
4. J.F. Marko, *Europhys. Lett.* **38**, 183 (1997).
5. R.D. Kamien, T.C. Lubensky, P. Nelson, C.S. O'Hern, *Europhys. Lett.* **38** (1997) 237-242.
6. C. Bouchiat, M. Mézard, preprint (cond-mat/9706050), (1997).
7. S.B. Smith, Y.J. Cui, C. Bustamante, *Science* **271**, 795-799 (1996).
8. M.D. Wang, H. Yin, R. Landick, J. Gelles, S.M. Block, *Biophys. J.* **72**, 1335-1346 (1997).
9. A. Stasiak, E. Di Capua, *Nature* **299**, 185-186 (1982).
10. T.R. Strick, J.F. Allemand, D. Bensimon, A. Bensimon, V. Croquette, *Science* **271**, 1835-1837 (1996).
11. P. Cluzel *et al.*, *Science* **271**, 792-794 (1996).
12. R.D. Kamien, T.C. Lubensky, P. Nelson, C.S. O'Hern, "Twist-Stretch Elasticity of DNA", *Statistical Mechanics in Physics and Biology, MRS Proceedings*, Vol. 463, MRS (Pittsburgh, 1997).
13. J.D. Moroz, P. Nelson, Torsional Directed Walks, Entropic Elasticity, and DNA Twist Stiffness, to appear in *Proc. Natl. Acad. Sci. USA* (1997).
14. The helical geometry of the DNA molecule alone cannot explain the twist-stretch coupling. Consider straightening out the helical sugar-phosphate backbones without changing the length per phosphate 0.6 nm or the distance from the center of the molecule 0.9 nm. Since each base pair step is $h = 0.34$ nm, the circumferential length per step is $l_c = \sqrt{0.6^2 - 0.34^2}$ nm. The corresponding twist angle per step is given by [15] $\theta = 2 \arcsin((l_c/2)/0.9 \text{ nm}) = 32^\circ$. An extension of the molecule by $\delta h/h = \epsilon$, yields an untwisting by an amount $\sigma = \delta\theta/\theta = \sigma_{\epsilon=0} - \epsilon/2.0$. The twist response to stretch given by the coefficient of ϵ in σ versus ϵ is much smaller than the coefficient (≈ 7) found experimentally [10].
15. C.R. Calladine, H.R. Drew, *Understanding DNA: the molecule and how it works* (Academic, London, 1992).
16. R. Lavery, H. Sklenar, *Journal Biomeolecular Structure & Dynamics* **6**, 655-667 (1989).
17. W. Saenger, *Principles of Nucleic Acid Structure* (Springer-Verlag, New York, 1984).
18. S. Krenk, *J. Appl. Mech.* **50**, 137-142 (1983).